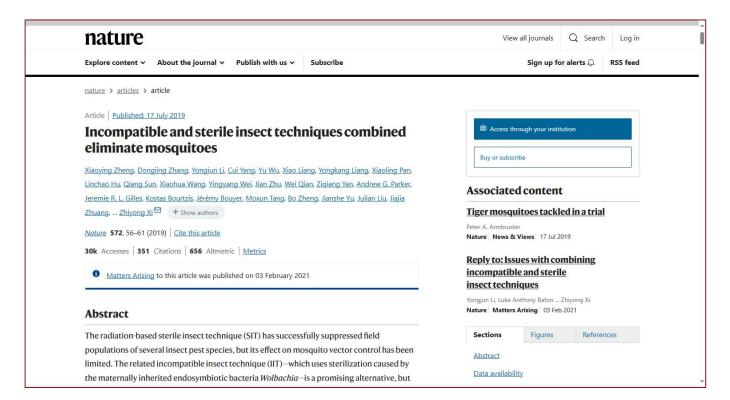


#### PCR Master Mix / DNA Polymerase



If wPip-positive larvae were observed, the sample was further screened by standard PCR using primers specific to the *ribosomal protein S6* (*rps6*) gene of *C. quinquefasciatus*, the only possible mosquito species with wPip co-occurring in the field sites, to exclude any false positives resulting from the collection of *Culex* larvae in the ovitraps. The specific-primers used for the assay were designed for *rps6* gene and consisted of: *rps6F*: 5'-TGCCGCGTCGTCTTGAATC-3'; and *rps6R*: 5'-GTATTGACCTCGTCGCGCTT-3'. The 20  $\mu$ l PCR reaction consisted of 2  $\mu$ l DNA template, 10  $\mu$ l PCR Master Mix (Dongsheng), 1  $\mu$ l of each primer (10  $\mu$ M) and 6  $\mu$ l ddH<sub>2</sub>O. The PCR conditions comprised of 5 min at 98 °C, followed by 40 cycles of 30 s at 98 °C, 5 s at 55 °C, 30 s at 72 °C, and then 10 min at 72 °C for the final extension. PCR products were electrophoresed on a 1.5% agarose gel, which contained 1  $\mu$ g/ml ethidium bromide. If a product size of approximately 350 bp was obtained, the sample was considered to contain *w*Pip derived from *Culex* mosquitoes.

Incompatible and sterile insect techniques combined eliminate mosquitoes.

Nature. (IF: 69.50)



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### PCR Master Mix / DNA Polymerase





Narrow mutational signatures drive acquisition of multidrug resistance in the fungal pathogen Candida Glabrata.

Current Biology. (IF: 10.90)

#### Target FKS and ERG3 sequencing

All ani-exposed samples (ANI, ANIFLZ and FinA) were examined for mutations in one region of FKS1 and two regions of FKS2 encompassing echinocandin resistance mutational HSs. Three samples without mutations in the above-mentioned HSs were also inspected in the HS2 of FKS1. All the new FKS mutations are in Data S2. We used PCR primers described earlier<sup>68</sup> (Table S5). ANI samples not subjected to WGS were also amplified by two PCRs with two sets of primers (Table S5) to obtain ERG3 sequences PCRs were carried out by using Taq DNA polymerase from DongShengBio. The reaction mixture included primers of concentration of 0.4 µM, 20 µL Tag DNA polymerase, 1 µL liquid sample grown for 24-48 h in YPD and water up to a final volume of 40 µL. Optimase ProtocolWriter was used to develop conditions for each primer set.

We tested for the possible trajectories of final FKS and ERG3 mutations in the 10 ANI samples subjected to WGS and presenting ERG3 alterations to infer which might have appeared first in the evolution. We selected and analyzed single colonies from our glycerols stocks of stored populations after the 2nd passage at 0.032, 0.064, 0.128 and 0.256 ug/ml ani (beginning of the adaptation). PCRs were carried out as described above.

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### PCR Master Mix / DNA Polymerase



#### 2.3. Detection of virulence genes

Nine virulence genes (vvhA, luxR, tdh, trh, toxR<sub>vc</sub>, flaC, ChiA, hlyA and vhpA) of Vibrio species were amplified by PCR. The primers are given in Table 1. The final volume of the amplification PCR was 50 μL, including 2 μL DNA template, 25 μL 2 × pfu Mix (Guangzhou Dongsheng Biotech, Guangzhou, China),  $2 \mu L$  forward and reverse primers, and  $19 \mu L$  ddH<sub>2</sub>O. PCR products were separated on a 2 % agarose gel by using 145 V/360 A, and the bands were recorded by an image analyzer BioDoc-ItTM system (UVP Inc., Upland, CA, USA).

Seasonal variation, virulence gene and antibiotic resistance of Vibrio in a semi-enclosed bay with mariculture (Dongshan Bay, Southern China). Marine Pollution Bulletin. (IF: 7.00)



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### PCR Master Mix / DNA Polymerase



the progression of SELEX, the amount of eluted ssDNA was measured by Nanodrop device (Thermo-Scientific, USA). An ssDNA library for the following round was prepared by an asymmetric PCR protocol using HS™ Mix (Dongsheng Biotech, Guangzhou, China, http://www.dongshengbio.com) with a molar ratio of primer-Fw/primer-Rv = 50:1. All the PCR amplifications were performed as follows: 1 cycle at 95 °C for 5 min; 30 cycles at 94 °C for 40 s, 50 °C for 30 s, 72 °C for 30 s; followed by 1 cycle at 72 °C for 10 min. The screening was repeated six times, and the last round eluted ssDNAs were amplified by standard PCR protocol. The

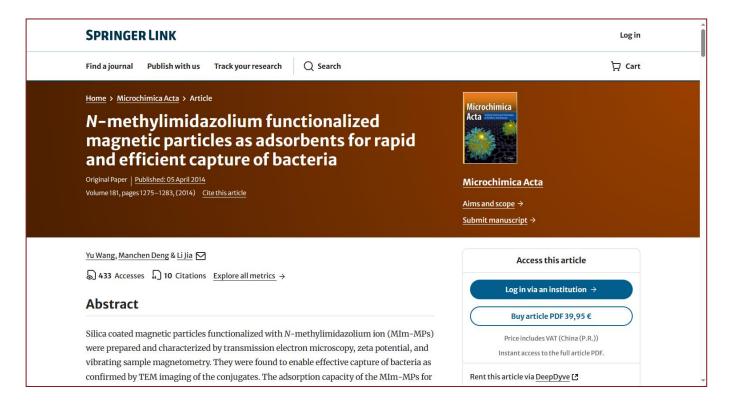
DNA aptamer for use in a fluorescent assay for the shrimp allergen tropomyosin. Microchimica Acta. (IF: 6.41)



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### PCR Master Mix / DNA Polymerase



A GeneRuler 100-bp DNA ladder was purchased from Fermentas (Beijing, China, www.fermentas.com) and consisted of ten fragments ranging from 100 to 1,000 bp with a total concentration of 0.5 mg mL-1. Taq DNA polymerase (5 U μL<sup>-1</sup>) was purchased from Dongsheng Biotech (Guangzhou, China, www.dongshengbio.com). 25 mM MgCl2, 10×PCR buffer (100 mM Tris-HCl [pH 8. 500 mM KCl), and deoxynucleotide triphosphate (dNTP) mixture (including dATP, dGTP, dCTP, and dTTP, where the concentration of each dNTP was 2.5 mM) were purchased

N-methylimidazolium functionalized magnetic particles as adsorbents for rapid and efficient capture of bacteria. Microchimica Acta. (IF: 6.41)



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### PCR Master Mix / DNA Polymerase



volume of 10ul, including 5ul 2xTaqMasterMix (DSBIO, China), 0.5ul each specific primer (10uM), 0.5ul DNA template and 3.5ul ddH<sub>2</sub>O. The PCR amplification reaction was carried out as following: pre-denaturation at 95 °C for 5 min; followed by 35 cycles of denaturation at 95 °C for 30 s, annealing at 58 °C for 30 s, and extension at 72 °C for 30s; and finally an extension step of 72 °C for 5 min. The amplified products were separated by 1.5% agarose gel. Then, another 16 DNAs (8 females

Screening and characterization of sex-specific markers developed by a simple NGS method in mandarin fish (Siniperca chuatsi).

Aquaculture. (IF: 5.14)



Web: www.gdsbio.cn



### PCR Master Mix / DNA Polymerase



Based on the filtrated X chromosome-specific sequences, primers were designed by Primer Premier 5.0 software (http://www.premierbio soft.com/). DNAs from XX, XY and YY individuals were used as templates to test the feasibility and universality of X chromosome-specific markers. Each PCR reaction was performed in a total volume of 10  $\mu$ l, including 5  $\mu$ l 2 × Tap MasterMix (DSBIO, China), 0.5  $\mu$ l each specific primer, 0.5  $\mu$ l DNA template and 3.5  $\mu$ l ddH<sub>2</sub>O. The PCR amplification reaction was carried out as following: pre-denaturation at 95 °C for 5 min; followed by 30 cycles of denaturation at 95 °C for 30 s, annealing at 60 °C for 30 s, and extension at 72 °C for 30s; and an extension step of 72 °C for 5 min finally. The amplified PCR products were visualized by gel electrophoresis on 1% agarose gels.

Screening and characterization of X chromosome-specific markers in mandarin fish (Siniperca chuatsi).

Aquaculture. (IF: 5.14)





### PCR Master Mix / DNA Polymerase



#### 2.4. Development and verification of male-specific sequences

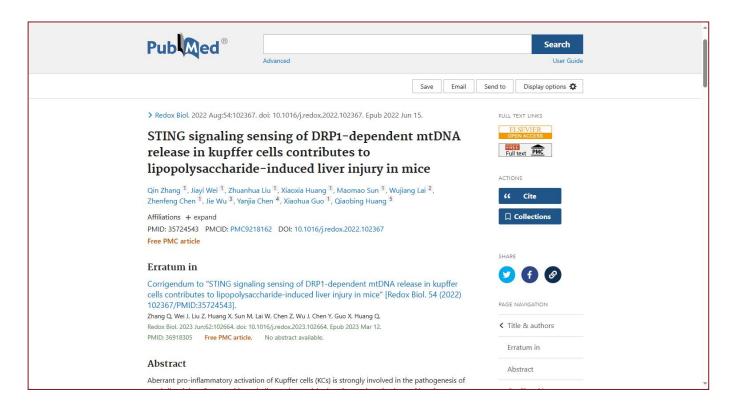
According to the obtained sex-specific sequences, a total of 20 primer pairs were designed using Primer 5.0. These primers were first verified in DNA samples of eight female and eight male S. hollandi. Then, the valid primers were further verified in DNA samples of another twenty four females and twenty four males. The PCR amplification reaction was carried out in a total of 10 µl reaction system, including 5 µl 2 x Taq Master Mix (GDSBIO, China), 0.5 µl each primer (10 uM), 0.5 µl DNA template and 3.5 µl ddH2O. The PCR amplification conditions include 5 min at 95 °C for pre-denaturation, following by 33 cycles 30 s at 95 °C, 30 s at 55 °C, 30 s at 72 °C, and a final extension for 5 min at 72 °C. Gel electrophoresis was used to detect the PCR products by 1.2% agarose gel.

Screening and characterization of sex-specific markers by NGS sequencing in Spinibarbus hollandi with implication of XY sex determination system. Aquaculture. (IF: 5.14)

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### **qPCR Master Mix**



#### 2.6. Quantitative real-time PCR

Total RNA from KCs was isolated with Trizol reagent, 500 ng of total RNA was reverse-transcribed into cDNA by using the RT-PCR Mix for qPCR kit (R1031, GDSBio) in which the genomic DNA was removed by DNase digestion. Real-time qPCR was conducted on the 7500 Real-Time PCR System (Applied Biosystems) with a Power Green qPCR Mix (P2102a, GDSBio) according to the manufacturer's procedures. The results of relative expression of mRNA of IFN- $\beta$ , TNF- $\alpha$  and IL-1 $\beta$  were normalized to the Ct value of GAPDH in each sample. The primers for qPCR analysis of sequences were presented in Supplementary Table 1.

STING signaling sensing of

DRP1-dependent mtDNA release in kupffer cells contributes to

lipopolysaccharide-induced liver injury in mice

Redox Biology. (IF: 10.79)





#### **qPCR** Master Mix



#### 2.7. Quantitative real-time polymerase chain reaction (qPCR)

Three liver samples were randomly selected from each treatment for total RNA isolation, cDNA synthesis, and quantitative real-time polymerase chain reaction (qPCR). qPCR was performed using a Roche LightCycler 96 (Roche, Basel, Switzerland). The total reaction system was 15  $\mu L$ , including 7.5  $\mu L$  of 2  $\times$  SYBR Premix Ex Taq II (GDSBio, Guangzhou, China), 0.3  $\mu L$  of each primer (10  $\mu M$ ), 2  $\mu L$  of cDNA, and 4.9  $\mu L$  of ddH2O. The reaction program adopted a threel-step method: pre-denaturation at 95 °C for 180 s, followed by 40 cycles of 10 s at 95 °C, 30 s at 60 °C, and 30 s at 72 °C. A dissociation curve analysis was performed at the end of amplification to determine target specificity. The relative expression levels were measured according to the threshold cycle value and normalized using the  $-\Delta \Delta Ct$  method. Rps4x was used as reference gene owing to its stable expression in liver samples under different treatment conditions. All primer sequences used for qPCR are listed in Table S1.

Long-term exposure to bisphenol A and its analogues alters the behavior of marine medaka (Oryzias melastigma) and causes hepatic injury.

Science of The Total Environment. (IF: 10.75)



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### **qPCR Master Mix**



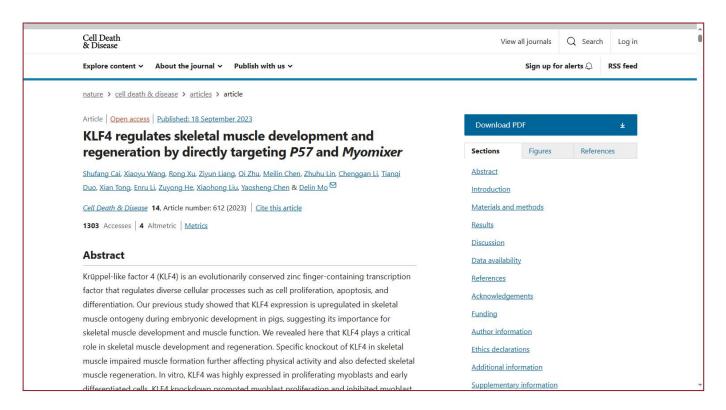
reaction test (RT-PCR) were purchased from Sangon Biotechnology Co., Ltd. (Shanghai, China). GSH was provided by Yuanye Bio-Technology Co., Ltd. (Shanghai, China). The 2',7'-dichlorodihydro-fluorescein diacetate (DCFH-DA), a cytosolic reactive oxygen species (ROS) probe, was purchased from Sigma-Aldrich Co., Ltd. (St. Louis, USA). SYBR green was purchased from Dongsheng Biotech Co., Ltd. (Guangzhou, China). The RevertAid First Strand cDNA synthesis kit and CellTracker deep red dye were from Thermo Fisher Scientific Co., Ltd. (MA, USA). Cell-tracker green CMFDA was from Maokang

CpG-Conjugated Silver Nanoparticles as a Multifunctional Nanomedicine to Promote Macrophage Efferocytosis and Repolarization for Atherosclerosis Therapy. ACS Applied Materials & Interfaces. (IF: 9.50)





#### **qPCR Master Mix**



#### RNA extraction and Real-time quantitative PCR

Total RNA was extracted from cultured C2C12 cells and regenerating TA muscles using Trizol Reagent (Invitrogen). Then, cDNA was synthesized from 1 µg total RNA using StarScript II First-strand cDNA Synthesis Mix (Genestar, Beijing, China). Real-time quantitative PCR (qPCR) analyses were performed on LightCycler 480 II (Roche, Basel, Switzerland) using SYBR Green qPCR Mix (GDSBio, Guangzhou, China), with GAPDH as an internal control for normalization. Primers are listed in Table S3.

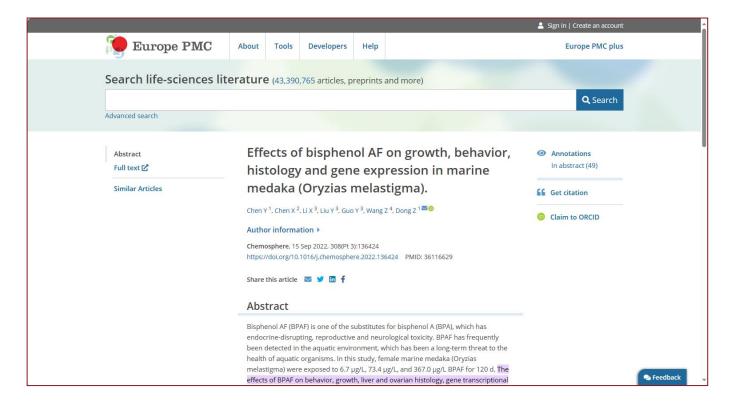
KLF4 regulates skeletal muscle development and regeneration by directly targeting P57 and Myomixer. Cell Deadth & Disease. (IF: 9.00)



Web: www.gdsbio.cn Tel: 020-87791356



### **qPCR Master Mix**



scriptional expression levels of target genes (Li et al., 2021). Quantitative polymerase chain reaction (qPCR) was performed using the SYBR® Green qPCR Mix (Dongsheng Biotech Co., Ltd., Guangzhou, China). All reactions were performed on the LightCycler® 96 PCR system (Roche, Basel, Switzerland). The list of primers is shown in Table S1. The PCR system (15  $\mu$ L) contained 7.5  $\mu$ L of 2 x SYBR® Green qPCR Mix<sup>a</sup>, 0.4  $\mu$ L of each forward and reverse primers (10  $\mu$ M), 1.5  $\mu$ L of cDNA samples, and 5.2  $\mu$ L of nuclease-free water. The qPCR reaction conditions were as

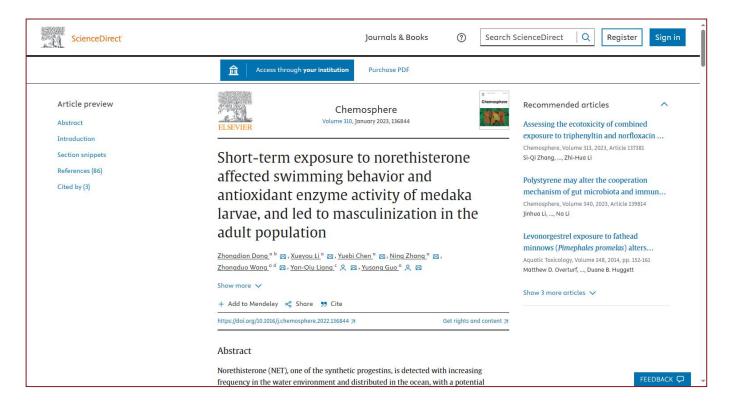
Effects of bisphenol AF on growth, behavior, histology and gene expression in marine medaka (Oryzias melastigma).

Chemosphere. (IF: 8.94)





### **qPCR Master Mix**



beta-2 (actb2) and glyceraldehyde-3-phosphate dehydrogenase (gadph) were used as internal reference genes (Dong et al., 2022). Roche LightCycler 96 (Roche, Basel, Switzerland) was used to perform the qPCR reaction, and the qPCR reaction reagent was Power Green qPCR Mix (GDSBio, Guangzhou, China). The system formulation and reaction procedure of qPCR refer to our previous study (Li et al., 2022). The relative mRNA expression level of target gene was transformed as Log2 according to the  $2^{-\Delta\Delta Ct}$  method (Livak and Schmittgen, 2001). Graph-Pad Prism 9 (GraphPad Software, Inc., La Jolla, CA, USA) was used to analyze and graph the data (expressed as Log2 form).

Short-term exposure to norethisterone affected swimming behavior and antioxidant enzyme activity of medaka larvae, and led to masculinization in the adult population.

Chemosphere. (IF: 8.94)

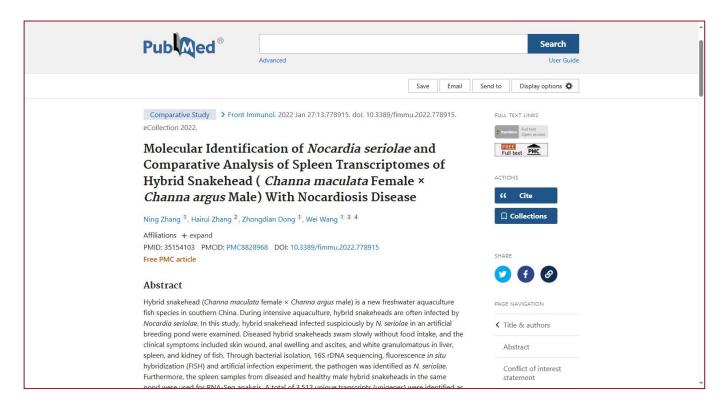


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#### **qPCR** Master Mix



#### qPCR Validation

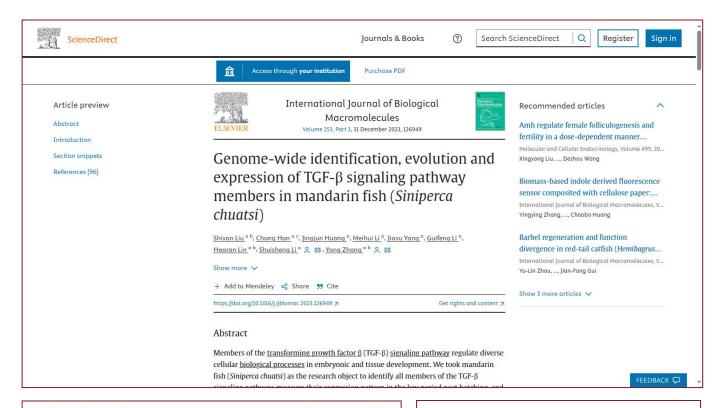
In order to validate the results of the RNA-Seq, twenty DEGs were randomly selected for validation by qPCR. Primers were designed according to the transcriptome sequences (**Supplementary Table 1**). qPCR was performed on Roche LightCycler 96 (Roche, Forrentrasse, Switzerland) using SYBR® Green qPCR Mix (GDSBio, Guangzhou, China). The reaction was carried out using a qPCR mixture of 15.0 μL, containing 7.5 μL 2 X SYBR® Green qPCR, 0.6 μL of each forward and reverse primer, 1.5 μL cDNA and 4.8 μL ddH<sub>2</sub>O. qPCR amplification was done as follows: 180 s at 95°C for pre-incubation, followed by 40 cycles at 95°C for 15 s, 60°C for 15 s and 72°C for 30 s. Dissociation and melting curves of qPCR products were performed, and results were analyzed. β-actin and ef2b were used as reference genes to determine relative expression (5). The transcriptional data were

Molecular Identification of Nocardia seriolae and Comparative Analysis of Spleen Transcriptomes of Hybrid Snakehead (Channa maculata Female × Channa argus Male) With Nocardiosis Disease. Frontiers in Immunology. (IF: 8.79)





#### **qPCR** Master Mix



According to the genome and transcriptome data of mandarin fish, the open reading frames (ORFs) of sex-associated genes (amh, amhr2, gdf9, bmp15, gsdf, gdf3, smad1, smad5, smad9, inha) were predicted, and gene-specific primers were designed using Primer 5.0 software (Table S1). Then, qRT–PCR was conducted using SYBR Green qPCR Mix (GDSBio, China) and performed on a Roche LightCycler 480 real-time PCR system. The qRT–PCR program was as follows: predenaturation at 95 °C for 3 min; followed by 45 cycles of 95 °C for 10 s, 58 °C for 20 s, and 72 °C for 15 s; and a final extension at 72 °C for 5 min, ending with a dissociation curve process. All qRT-PCRs used  $\beta$ -actin as the internal control and were carried out in triplicate to confirm the results. The specificity of each primer amplification was conducted with the dissociation curve. Under a fivefold cDNA dilution series, the qPCR

Genome-wide identification, evolution and expression of TGF- $\beta$  signaling pathway members in mandarin fish (Siniperca chuatsi).

International Journal of Biological Macromolecules. (IF: 8.20)



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#### **qPCR Master Mix**



Total RNA was extracted from all obtained tissues using a Trizol RNA extraction kit (Invitrogen, USA) following the manufacturer's instructions. The quantity and quality of RNA were further detected using Nanodrop 2000c (ThermoFisher, USA). Then, all cDNA templates were synthesized using RT-PCR Mix for qPCR (GDSBIO, China) and then stored at -20 °C.

To detect expression profiles of SbTLRs, quantitative real time PCR (qRT-PCR) reaction was performed using the SYBR Green qPCR Mix (GDSBIO, China) on a LightCycle 480 System (Roche, Germany). All primers used in qRT-PCR reaction were designed by Primer 3.0 (Table 1) and further validated using agarose gel electrophoresis and Sanger sequencing. All qRT-PCR reactions were performed in a volume of 10 μL, containing 0.5 µL cDNA, 0.5 µL of each primer (10 µM), 5 µL SYBR Green qPCR Mix, and 3.5 μL of ddH2O. The PCR cycling conditions are pre-

Molecular characterization and expression analysis of nine toll like receptor (TLR) genes in Scortum barcoo under Streptococcus agalactiae infection.

International Journal of Biological Macromolecules. (IF: 8.20)



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### **qPCR Master Mix**



#### Quantitative RT-PCR analysis

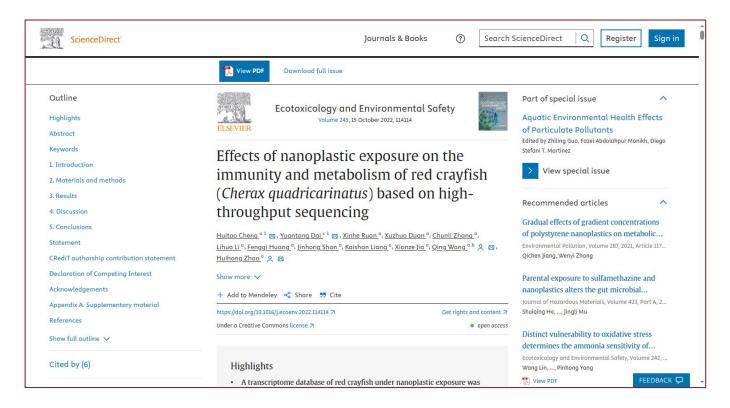
Total cellular RNAs were extracted using Trizol reagent (Ambion). cDNAs were synthesized from 2µg total RNA using oligo(dT) primers and M-MLV reverse transcriptase system (Promega). cDNA was used as template for quantitative PCR using a Bio-Rad CFX-96 real time PCR apparatus and SYBR green master mix (Dongsheng Biotech). PCR conditions were as follow: an initial denaturation at 94°C for 3 min, 40 cycles of 94°C for 15 s, 60°C for 15 s and 72°C for 20 s. The specificity of the amplified PCR products was confirmed by melting curve analysis after each reaction. The primers used were for human IFN-β, 5'-GCTTGGATTC

Inhibition of anti-viral stress granule formation by coronavirus endoribonuclease nsp15 ensures efficient virus replication.
Plos Pathogens. (IF: 7.46)





### **qPCR Master Mix**



#### 2.3.4. Verification of DEGs by quantitative real-time PCR (qRT-PCR)

To verify the reliability of the transcriptome data, 12 differentially expressed genes were selected for qRT-PCR validation. Based on the 12 genes and  $\beta$ -actin sequences, Premier6 software were used to design specific primers for qRT-PCR analysis (Table S1). The PCR reaction system was 10  $\mu L$  consisting of 5  $\mu L$  2  $\times$  SYBR Ex Taq (Guangzhou Dongsheng Biotechnology Co., Ltd.), 0.2  $\mu L$  of each gene-specific primer (10 nmol), 1  $\mu L$  cDNA and 3.6  $\mu L$  ribonuclease-free water composition. Cycling conditions were 3 min at 94 °C; 40 cycles of 94 °C for 15 s, 60 °C for 15 s, and 72 °C for 20 s. The relative expression was calculated by the  $2^{-\Delta\Delta CT}$  method with  $\beta$ -actin gene as the internal reference gene.

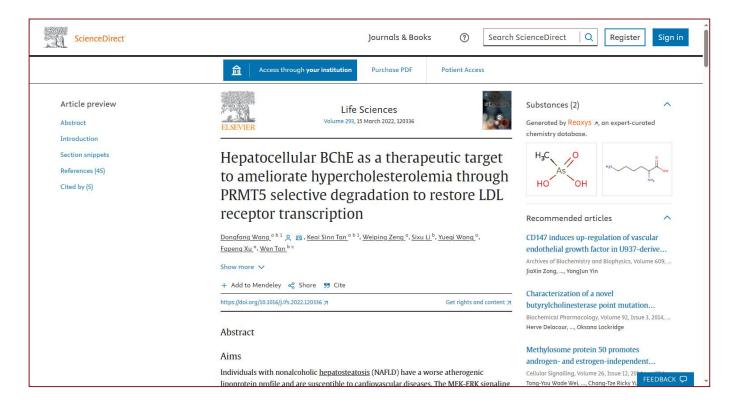
Effects of nanoplastic exposure on the immunity and metabolism of red crayfish (Cherax quadricarinatus) based on high-throughput sequencing.

Ecotoxicology and Environmental Safety. (IF: 7.13)





#### **qPCR Master Mix**



Total RNA was extracted from Aml-12 cells using TRIzol Reagent (Invitrogen, 15596026) as described by the manufacturer. RNA integrity was determined by agrose eletrophorosis, and RNA concentration was measured by NanoDrop 2000. Reverse transcription was performed to synthesize single-stranded cDNA using 5  $\mu$ g RNA as the template according the protocol of the manufacturer (Hifair® III 1st Strand cDNA Synthesis Kit (gDNA digester plus), 11139ES60, Yeasen Biotechnology). Real-time PCR was performed with Power Green qPCR Mix No ROX (P2104, GDSbio, China) on the Roche LightCycler® 96 Instrument.  $\beta$ -actin was used as the internal control to normalize the data. The amplification program was as follows, 95 °C for 3 min, and 40 cycles at 95 °C for 15 s, 60 °C for 15 s, 72 °C for 20 s. Dissociation curves were analyzed at the end of the amplification. The fold change of gene

Hepatocellular BChE as a therapeutic target to ameliorate hypercholesterolemia through PRMT5 selective degradation. Life Sciences. (IF: 6.78)





### **qPCR Master Mix**



The PCRs were performed in 20  $\mu$ L reaction volumes containing  $2\times$  SYBR Green Mix 10  $\mu$ L, Primer Mix 1  $\mu$ L 9 (DongshengBio, Guangzhou, China), DNA template 1  $\mu$ L, ddH<sub>2</sub>O 8  $\mu$ L. Denaturation program (95 °C for 2 min), amplification and quantification program repeated 40 times (95 °C for 15 s, 59 °C for 20 s, 72 °C for 20 s with a single fluorescence measurement); Melting curve program (60–95 °C

Bacterial Cellulose Membranes Used as Artificial Substitutes for Dural Defection in Rabbits.

International Journal of Molecular Sciences. (IF: 6.21)





### **qPCR Master Mix**



Continued			
REAGENT or RESOURCE	SOURCE	IDENTIFIER	
FerroOrange	DoJindo	F374	
SYBR Green qPCR Mix	GDSBio	P2092	
Liperfluo	DoJindo	L248	

Newcastle-disease-virus-induced ferroptosis through nutrient deprivation and ferritinophagy in tumor cells. iScience. (IF: 6.11)

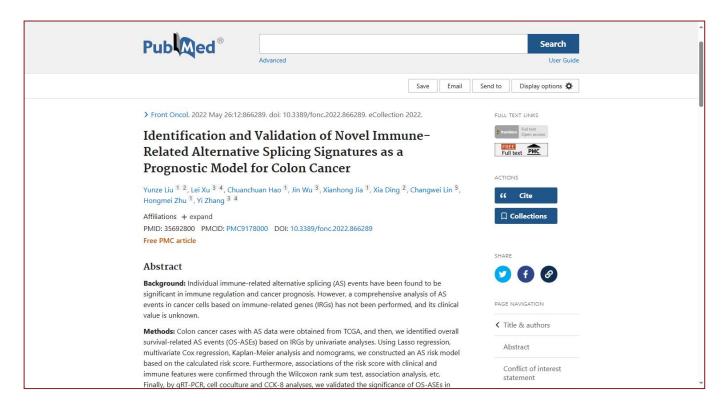


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#### **qPCR** Master Mix



#### RNA Extraction and gRT-PCR

Total RNA was extracted from cells and tissues using the TRIzol method (Dongsheng Bio. #R1022) following the protocol. Then, the obtained RNAs were processed for cDNA synthesis. qRT-PCR was then performed using SYBR Green qPCR Mix (Dongsheng Bio. #P2092) and analysed on a Roche LightCycler system. The expression levels of the target genes were normalized based on the expression level of GAPDH. The primer sequences used for amplification and siRNA sequences are listed in the Supplemental Material.

Identification and Validation of Novel Immune-Related Alternative Splicing Signatures as a Prognostic Model for Colon Cancer.

Frontiers in Oncology. (IF: 5.74)



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#### **qPCR Master Mix**



#### 2.7. Quantitative PCR (qPCR)

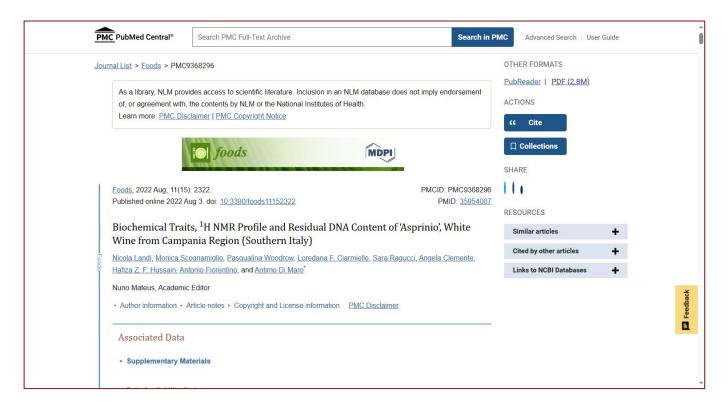
RNA was isolated by a TRIzol Trizol reagent, following the manufacturer's instructions. The cDNA was synthesized via the reverse transcription (Accurate Biology, AG11706). SYBR Green (GDSBio, P2105) was used to perform qPCR using the Real-Time PCR instrument (Bio-Rad). The data were calculated though 2<sup>-ΔΔCt</sup> method to compare the difference. The following primers were used to amplify the target genes — MMP-9 forward: 5'-CGCCTCTGGAGGTTCGAC-3', reverse: 5'-AACT-CACGCGCCAGTAGAAG-3'; VEGF forward: 5'-ATCTTCAAGC-CATCCTGTGTGC-3'. reverse: 5'-CAAGGCCCACAGGGATTTTC-3'; TGF-β1 forward: 5'-CGACTCGCCAGAGTGGTTAT-3'. reverse: 5'-CGGTAGT-GAACCCGTTGATGT-3'; GAPDH forward: 5'-CCTCCTTCTGCACA-CATTTGAA-3', reverse: 5'-GAAGATGGTGATGGGATTTC-3'.

Blockade of CBX4-mediated  $\beta$ -catenin SUMOylation attenuates airway epithelial barrier dysfunction in asthma. International Immunopharmacology. (IF: 5.71)





### **qPCR Master Mix**



#### 2.7.2. DNA Grapevine and Yeast Quantification

DNA quantification was carried out by quantitative PCR analyses using SYBR Green qPCR Mix (high ROX; GDSBio) by Applied Biosystems QuantStudio 5 Real-Time PCR System (Thermo Fisher Scientific, Rodano (MI), Italy). The reaction mix (20  $\mu$ L) consisted of: 2 × SYBR Green qPCR Mix, Forward 10  $\mu$ M for each primer, and DNA template 50 ng. The amplification protocol was: 95 °C initial denaturation step for 3 min, followed by amplification cycles (40×) of denaturing at 95 °C for 10 s, annealing at 60 °C for 10 s and extension at 72 °C for 20 s. Amplification products were visualized on 1.5% (w/v) agarose gel, using a UV light. Primers used in this study are listed in Table S2.

Biochemical Traits, 1H NMR Profile and Residual DNA Content of 'Asprinio', White Wine from Campania Region (Southern Italy). Foods. (IF: 5.56)





### **qPCR Master Mix**



#### 2.18. Quantitative real-time polymerase chain reaction (qRT-PCR)

qRT-PCR was performed using QuantStudio 6 Flex real-time PCR system (Life Technologies, Carlsbad, CA, USA) together with the Power Green qPCR Mix (Guangzhou Dongsheng Biotech Co., Ltd., China). Primers synthesized by the Xiamen Borui Biotechnology co., LTD (China) were listed in Supplementary Material 1. gyrB was used to normalize the gene expression levels. The relative levels of gene expression were calculated by  $2^{-\Delta\Delta Ct}$  method (Luo et al., 2016).

Vvrr2: A new Vibrio ncRNA involved in dynamic synthesis of multiple biofilm matrix exopolusaccharides, biofilm structuring and virulence.

Aquaculture. (IF: 5.14)



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### **qPCR Master Mix**



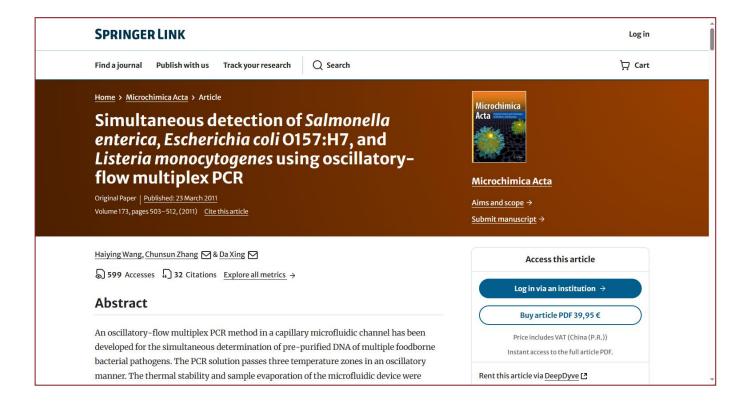
RT-qPCR. Total RNA was extracted from the cell samples using traditional extraction methods and reverse transcribed using M-MLV reverse transcriptase (Promega; M1705). The RT-qPCR probes were obtained using SYBR green qPCR mix (GDSBio, Guangzhou Gongsheng Biotech Co., Ltd.; P2093). The primers targeting different genes were as follows: c-Myc (35), SHMT1 (44), MTHFD1L (45), and MTHFD2 (46). The NDV NP primer sequences were as follows: 5'-CAACAATAGGAGTGGAGTGTCTGA-3' and downstream primer, 5'-CAGGGTATCG GTGATGTCTTCT-3'. All primer pairs were synthesized by Shanghai Sangon Biotech (Shanghai, China). All samples were standardized according to the mRNA level of β-actin.

Newcastle Disease Virus Manipulates Mitochondrial MTHFD2-Mediated Nucleotide Metabolism for Virus Replication.

Journal of Virology. (IF: 5.10)







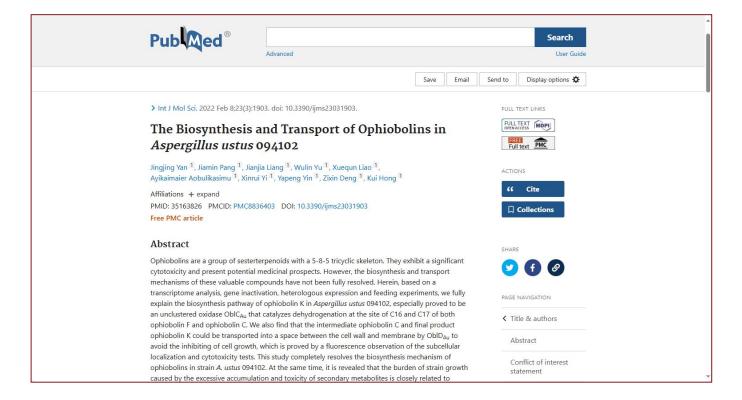
The DNA markers, which contain 500, 400, 300, 250, 200, 150, 100 and 50 bp DNA fragments, were from Dongsheng Biotech Co., Ltd. (Guangzhou, China, www.dongshengbio.com).

Simultaneous detection of Salmonella enterica, Escherichia coli O157: H7, and Listeria monocytogenes using oscillatory-flow multiplex PCR.

Microchimica Acta. (IF: 6.41)







#### 4.1. General Experiential Materials

DNA fragments for construction of plasmids were amplified using I-5<sup>TM</sup> 2×High-Fidelity Master Mix (TSINGKE Biotech, Beijing, China), whereas PCR for screening of transformants was performed using 2×Es Taq MasterMix (CWBIO, Beijing, China). Primers were synthesized by TSINGKE Biotech (Beijing, China) and sequencing was carried out in TSINGKE Biotech (Beijing, China). The 1 kb ladder (GDSBio, Guangzhou, China) was used as DNA marker. All restriction endonucleases were purchased from New England BioLabs (NEB; Ipswich, MA, USA). T-Vector pMD19 (Simple) was purchased from TaKaRa (Kyoto, Japan). ClonExpress Ultra One Step Clone Kit (Vazyme, Nanjing, China) was used for construction of plasmids rapidly.

The Biosynthesis and Transport of Ophiobolins in Aspergillus ustus 094102.

International Journal of Molecular Sciences. (IF: 6.21)



Web: www.gdsbio.cn





**Figure 1.** Gel electrophoresis of PCR products for *E. coli*-specific primers *uidA* (a) marker genes ( $bla_{TEM}$ ,  $bla_{CTX-M}$ , and  $bla_{SHV}$ ) (**b-d**) and five virulence factors marker genes (pai, hly, cnf-1, sfa and afa) (**c-f**). The amplified DNA fragments produce of various *E. coli* strains with these marker primers: species-specific primer uidA (lanes 1–5 for strains 1–5) (a), CTX-M (lanes 1, 2 and 3 for control negative and strains 4 and 36, respectively) (b), TEM (lanes 1 and 2 for strains 4 and 36), hly (lanes 3 and 4 for strains 3 and 4 and lane 5 for control negative), and sfa (lanes 7 and 8, for strains 3 and 4) (c), SHV (lanes 1 and 2, for strains control positive and S36), afa (lanes 3 and 4, for strains 2 and 3) (d), cnf-1 (lanes 1, 2, 3, 4, 5 and 6 for strains 4, 5, 6, 7, 8 and control-ve) (e), and pai (lanes 1, 2, and 3, for strains 4, 6 and 7) (f), and lane  $\frac{100}{2}$  bp Ladder (GDSBio Marker) (b-e) and 300 bp (GeneDireX, Marker) as shown in (a,f). A 7 μL of the PCR products and ladder were pipetted into a prepared 1.5% agarose gel stained with 5 μL of Safe Gel Stain Dye. Key: pai; pathogenicity island; hly; hemolysin, sfa; S-fimbrial adhesion, cnf-1; cytotoxic necrotizing factor-1, and afa; a fimbrial adhesion and S = strain.

Molecular Characterization of Extended Spectrum β-Lactamase (ESBL) and Virulence Gene-Factors in Uropathogenic Escherichia coli (UPEC) in Children in Duhok City, Kurdistan Region, Iraq.

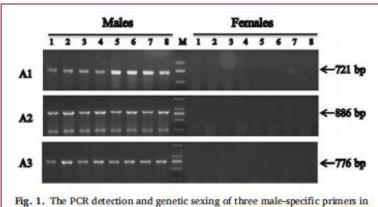
Antibiotics-Basel. (IF: 5.22)



Web: www.gdsbio.cn







eight females and eight males of S. hollandi. The DL 2000 DNA marker (GDSbio, China) is shown in the middle.

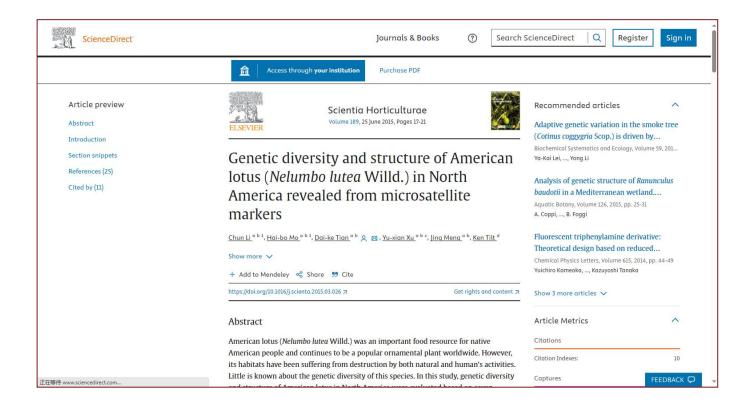
Screening and characterization of sex-specific markers by NGS sequencing in Spinibarbus hollandi with implication of XY sex determination system.

Aquaculture. (IF: 5.14)



广州东盛生物科技有限公司





#### Assessing genetic diversity of populations of topmouth culter

https://www.sciencedirect.com/science/article/pii/S0305197807000968

Clear and unambiguous bands in length ranging from 50 to 1200 bp were considered as usable, and the band sizes were estimated using a standard 100 bp DNA ladder (Dongsheng Biotech). T...

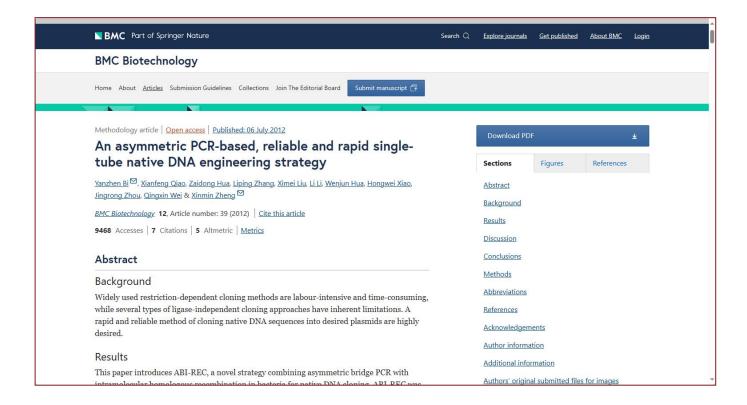
Genetic diversity and structure of American lotus (Nelumbo lutea Willd.) in North America revealed from microsatellite markers.

Scientia Horticulturae. (IF: 4.3)



Web: www.gdsbio.cn





#### **Enzymes and reagents**

ApaI, SalI, DpnI, BglII and XbaI restriction nucleases, *Taq* DNA polymerase and dNTP were purchased from Fermentas (Lithuania). KOD Plus high-fidelity DNA polymerase was purchased from Toyobo (Japan). A 1kb DNA ladder was from Dongsheng Co. Ltd. (Guangzhou, China). Enzymatic reactions were carried out under recommended conditions. All other chemicals used in the study were of molecular biology grade.

An asymmetric PCR-based, reliable and rapid single-tube native DNA engineering strategy.

BMC Biotechnology. (IF: 3.50)







purchased from Tiangen Co (Beijing, China). The 20bp ladder was purchased from Dongsheng Bio-tech Inc. (Guangzhou, China).

Cloning and identification of novel miRNAs in the flower organs of Korla fragrant pear at anthesis. Journal of horticultural science & biotechnology. (IF: 1.92)

